

Chemoenzymatic Asymmetric Total Synthesis of Phosphodiesterase Inhibitors: Preparation of a Polycyclic Pyrazolo[3,4-d]pyrimidine from an Acylnitroso Diels-Alder Cycloadduct-Derived Aminocyclopentenol

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Received September 9, 2004



Enzymatic resolution of Boc-protected 4-aminocyclopenten-1-ol **4c** gave both enantiomers **5c** and **6c** in high ee. Boc removal and separate condensation with chloropyrazolopyrimidine **18** provided elaborated 1,4-aminocyclopentenol derivatives **20** and **26**, respectively. Separate treatment of **20** and **26** with Pd(0) under basic conditions induced cyclization to unsaturated polycycles **22** and **27**, which, upon catalytic hydrogenation, were transformed to new cyclopentane-containing pyrazolopyrimidines **24** and **28**, analogues of recently described novel phosphodiesterase inhibitors.

Functionalized cyclopentane derivatives are major constituents of many natural products and analogues,¹ and stereoselective syntheses of cyclopentane derivatives have been especially important for the preparation of a number of carbocyclic nucleosides.² Functionalized aminocyclopentenol derivatives are also potential aminoglycosidase inhibitors.^{1c,3} We have previously described an efficient enzymatic resolution of 4-aminocyclopentol-1ol **4**⁴ derived from acylnitroso Diels-Alder adducts **3**⁵ (Scheme 1).

Using this process, both enantiomers of the Boc-protected 4-aminocyclopenten-1-ol **4** are available in optically pure form by employing different enzymes. The synthetic utility of related aminocyclopentenol derivatives has been

SCHEME 1



highlighted by several recent applications.^{6a,b,7} Derivatives of acetate 6 are ideally suited for the introduction of nucleophiles via palladium-mediated π -allyl chemistry.⁸ Here we report an application of enantiomerically pure aminocyclopentenol 5 or 6 in the synthesis of phosphodiesterase inhibitor analogues of recent interest. Phosphodiesterases (PDE) are involved in cyclic nucleotide regulation,⁹ and PDE inhibition as a target for therapeutic intervention is of considerable interest.¹⁰ More specifically, the beneficial effects of inhibition of the cGMP PDE for the treatment of cardiovascular diseases have been notable.¹¹ These biological properties resulted in numerous research programs directed toward the synthesis of PDE inhibitors and analogues thereof.¹² Polycyclic guanine derivatives of the general type 7 (Figure 1) were found to be potent inhibitors of PDE1 and PDE5 in vitro and potent antihypertensive agents in vivo.¹³ Polyclic pyrazolo[3,4-d] analogues of the type 8 and 9 have also been found to have similar activity.¹⁴ As

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10.1021/jo0484070 CCC: \$30.25 © 2005 American Chemical Society Published on Web 03/04/2005

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FIGURE 1. Polycyclic guanine PDE inhibitors **7** and pyrazolo-[3,4-*d*]pyrimidine analogues **8** and **9**.

SCHEME 2



shown in Scheme 2, we anticipated that related analogues 10 could be prepared by Pd(0)-mediated cyclizations of 12 to 11 followed by the reduction of the remaining double bond. By using enzymatically resolved compound 4, both enantiomers of 10 could be prepared by similar methods.

As indicated in Scheme 1, our previous publication reported the enzymatic resolution of a number of Nprotected aminocyclopentenol 4 to provide either 5 or 6 in high ee.⁴ The starting material **4** for this resolution was prepared through protection of hydroxylamine, oxidation, and trapping of the transient nitroso intermediate with cyclopentadiene to produce (\pm) -3 and N-O bond reduction using substoichiometric (20 mol %) Mo-(CO)₆ and NaBH₄ as we have also previously described.⁶ Although this method provided access to large amounts of optically pure **5a** and **6a**, further elaboration of these building blocks for the synthesis of complex carbocyclic nucleosides and other highly functionalized cyclopentanecontaining compounds of interest often required the exchange of the N-acetyl protecting group for one that could be removed under milder conditions. Therefore, we sought to improve the resolution of Boc-protected aminocyclopentenol (\pm) -4c which, using our reported methodology,⁴ gave acetate (-)-6c in 81% ee (without recrystallization). To develop a method that would also enable us to recycle the enzyme, immobilized Candida antarctica B^{15} was used. Racemic aminocyclopentenol (±)-4c was mixed with vinyl acetate and enzyme in wet CH₂Cl₂ or heptane/CH₂Cl₂ for 3–4 h at room temperature providing 6c in 40-50% yield and leaving 5c unreacted. After chro-

SCHEME 3



matographic separation, (-)-**6c** was determined to be significantly optically enriched, the ee was improved from 81% to >90% with the use of the resin-bound enzyme (ee was determined by hydrolysis of the acetate and acylation to give the corresponding Mosher ester as described earlier⁴). A single recrystallization of **6c** provided multigram amounts of (-)-**6c** in >98% ee. The chromatographically separated alcohol was enriched in **5c** and was isolated in 35-45% yield. After a single recrystallization, the compound was determined to be essentially optically pure (>99% ee) by Mosher ester analysis.

Pyrazole 16 was prepared via a modified procedure (Scheme 3).¹⁴ Condensation of ethyl cyanoacetate 14 with triethyl orthoformate generated ethyl (ethoxyethylene)-cyanoacetate 15, which was then converted to pyrazole 16 by treatment with benzylhydrazine. Condensation of 16 with phenyl isocyanate followed by base-promoted cyclization formed intermediate 17. Compound 17 was then converted to the desired chloride 18 by treatment with phosphorus oxychloride.

Next, as shown in Scheme 4, enantiopure 5c was converted to acetate 19. The Boc group was removed and the resultant TFA salt of 19 was neutralized and condensed with chloride 18 in the presence of triethylamine to afford 20 in 96% yield. Our next task was to perform the crucial palladium(0)-mediated cyclization reaction. Gratifyingly, treatment of 20 with in situ generated palladium(0) in THF induced formation of the desired product, 22, under basic conditions (Table 1). It was found that this reaction was sensitive to the base used. When the relatively weak base NEt₃ was used, the reaction proceeded slowly, and after overnight, only 20% of the starting material was consumed. When NaH was used, the reaction was complete in 3 h and gave the desired product in 79% yield. When the organic base 2-tert-butylimino-2-diethylamino-1,3-dimethylperhydro-1,3,2-diazaphosphorine, **21**, was utilized, the cyclization reaction gave the highest yield (94%) of product 22. Compound 22 was then subjected to catalytic hydrogenation. It was surprising to find that upon complete reaction of 22, rather than the desired hydrogenated product 24, the ring-opened product 23 was generated in 50% yield as the only isolable product after the reaction was carried out with Pd/C and H_2 . The structure of 23 was first determined by extensive 1D and 2D NMR analysis. It was further confirmed by synthesizing 23 through the simple coupling reaction of cyclopentylamine 25 with chloride 18 under conditions similar to those used to prepare 20 (Scheme 5).

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SCHEME 4



 TABLE 1. Pd(0)-Catalyzed Cyclization Reaction of 20 to

 22 under Basic Conditions

base	time (t)	yield of 22 (%)
$egin{array}{c} { m NEt}_3 \ { m NaH} \ { m 21} \end{array}$	overnight 3 h 3 h	10 (20% conversion) 79 94

SCHEME 5



TABLE 2. Hydrogenation Reaction of Compound 22under Different Conditions



The hydrogenation reaction was carried out under different conditions, which are summarized in Table 2. The desired selective reduction was finally achieved by the reaction of **22** with diimide¹⁶ (HNNH, generated in situ by reaction of aqueous hydrogen peroxide and hydrazine in EtOH solution) to give **24** in 50–60% yield. Starting with enantiomerically pure compound **6c**, the enantiomer of **24** also has been synthesized (Scheme 6).

Preliminary determination of phosphodiesterase (PDE1) inhibition at a concentration of 100 μ M revealed modest activity of 26% inhibition by precursor **20** and a more significant 71% inhibition by ring-opened product **23**. Both enantiomers of **24** and **28** showed modest PDE1 inhibition, with IC₅₀ values of 55.8 and 41.2 μ M, respectively.

In conclusion, we were able to develop a mild and potentially general protocol for the construction of poly-

SCHEME 6



cylic pyrazolo[3,4-*d*]pyrimidines in a highly efficient manner (54% overall yield starting from optically pure Boc-protected aminocyclopentenol 5c and 52% overall yield from 6c). In addition to the high chemical efficiency, this route should be compatible with peripheral variation that will allow access to a library of compounds suitable for biological study.

Experimental Section

Enzymatic Acetylations. To a solution of 4c (10 g, 50.25 mmol) in a mixture of wet heptane/dichloromethane (2.8:1, 380 mL) was added vinyl acetate (22.2 mL, 240.9 mmol). The wet heptane was prepared by thoroughly mixing with water in a 1 L bottle, the two layers were allowed to separate, and the organic layer was decanted and used. Resin-bound C. antarctica B lipase $(CAB^*)^{15}$ (1 g) was added, and the resultant mixture was shaken vigorously in an incubator-shaker at room temperature while the reaction was monitored by TLC. The conversion was complete within 3 h, although longer times had no deleterious effect. The mixture was filtered through a filter paper to remove the resin-bound enzyme. The enzyme was recovered and could be used a second time of resolution of 4c. The solvent was removed from the filtrate, and the mixture was then purified using column chromatography (hexanes/EtOAc 10:1) to afford 5.2 g (43%) of 6c as a white solid (90% ee, determined by hydrolysis of the acetate (K₂CO₃/MeOH) and derivatization with (S)-(+)- α -methoxy- α -(trifluoromethyl)phenyl acetyl chloride for Mosher ester analysis) The product, 6c, was recrystallized using hexanes to afford 4.6 g of 6c as a white, crystalline solid (98% ee): $[\alpha]_{\rm D} = -22.6 \ (c = 0.40, \text{ CHCl}_3); \text{ mp} = 57-58 \ ^{\circ}\text{C}; ^{1}\text{H NMR}$ (300 MHz, DMSO- d_6) δ 1.37 (s, 9H), 1.45 (dt, J = 13.5, 6.0 Hz, 1H), 1.98 (s, 3H), 2.68 (dt, J = 13.5, 7.5 Hz, 1H), 4.38 (m, 1H), 5.40 (m, 1H), 5.81 (dt, J = 5.4, 2.1 Hz, 1H), 5.90 (m, 1H), 7.11 (d, J = 7.5 Hz, 1H); ¹³C NMR (75 MHz, DMSO- d_6) δ 20.8, 28.2, 37.6, 53.3, 77.1, 77.8, 130.9, 137.3, 154.9, 170.1; HRMS [MH+] calcd for C₁₂H₁₉NO₄ 242.1392, found 242.1382. The unreacted alcohol 5c was recovered in 42% yield with 98% ee after recrystallization from hexanes/EtOAc at 0 °C: $[\alpha]_D = -69.0$

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 $(c=1.0, \text{CHCl}_3); \text{mp} = 103-104 \text{ °C}.$ Spectral data were identical to those previously reported.⁴

1-Benzyl-6-chloro-5-phenyl-1,5-dihydropyrazolo[3,4-d]pyrimidin-4-one (18). Compound 17 (1.1 g, 3.45 mmol)¹⁴ was heated with POCl₃ (16 mL) at reflux for 48 h. The solution was cooled and diluted with CH2Cl2. The organic layer was washed with saturated NaHCO₃ (Caution, vigorous exotherm and gas evolution), H₂O, and brine and dried with MgSO₄. After filtration and concentration, the residue was purified by column chromotagraphy eluting with hexanes/EtOAc from 10:1 to 5:1 to 3:1 to give 0.51 g (44% yield) of a white solid: mp = 159-161 °C; R_f = 0.65 (hexanes/EtOAc = 1:1); ¹H (300 MHz, CDCl₃) δ 5.49 (s, 2H), 7.21-7.23 (m, 2H), 7.25-7.39 (m, 5H), 7.48-7.55 (m, 3H), 8.07 (s, 1H); ¹³C (75 MHz, CDCl₃) δ 51.2, 104.3, 128.1, 128.1, 128.3, 128.7, 129.6, 129.6, 135.6, 136.0, 137.2, 148.3, 149.7, 157.2; IR (KBr) 3065, 1723, 1565, 1489, 1284, 1224, 1186, 1031, 712 cm⁻¹; HRMS (FAB) calcd for $C_{18}H_{14}N_4O^{35}Cl (M + H)^+ 337.0856$, found 337.0850.

Acetic Acid 4-(1-Benzyl-4-oxo-5-phenyl-4,5-dihydro-1Hpyrazolo[3,4-d]pyrimidinyl-6-amino)cyclopent-2-enyl Ester (20). To an ice-cold solution of 19 (50 mg, 0.21 mmol) in CH₂Cl₂ (2 mL) was added trifluoroacetic acid (0.17 mL). The mixture was stirred at 0 °C for 20 min, warmed to room temperature, and stirred for another 1 h. The solvent was removed by coevaporation with toluene $(2 \times 2 \text{ mL})$. The residue was dissolved in n-BuOH (2 mL). Chloride 18 (142 mg, 0.42 mmol) was added to the solution followed by Et₃N (0.26 mL, 1.89 mmol). The mixture was heated at 105 °C for 28 h under an Ar atmosphere. After the reaction was cooled, the solvent was removed and the residue was purified by column chromatography eluting with hexanes/EtOAc (3:1 to 1:1) to afford 89 mg (96% yield) of a white foam: mp = 64 °C; $R_f = 0.65$ (1:1 of hexanes/ EtOAc); $[\alpha]_{\rm D} = -7.2 \ (c = 1.0, \text{ CHCl}_3); \ ^1\text{H} (500 \text{ MHz}, \text{ CDCl}_3) \ \delta$ 1.44 (dt, J = 4, 14 Hz, 1H), 1.93 (s, 3H), 2.81 (q, J = 7.5 Hz, 1H), 4.23 (d, J = 7.5 Hz, 1H), 5.03–5.05 (m, 1H), 5.39 (s, 2H), 5.50-5.51 (m, 1H), 5.94-5.98 (m, 2H), 7.23-7.38 (m, 6H), 7.50-7.58 (m, 4H), 7.97 (s, 1H); ¹³C (125 MHz, CDCl₃) δ 21.0, 38.5, 50.6, 55.6, 77.2, 100.2, 127.8, 128.1, 128.6, 129.0, 129.1, 129.9, 130.6, 130.6, 133.2, 134.5, 135.9, 136.0, 136.7, 151.9, 152.4, 158.1, 170.3; IR (CH₂Cl₂, NaCl) v 2987, 1765, 1665, 1221, 880 cm⁻¹; HRMS (FAB) calcd for $C_{25}H_{24}N_5O_3$ (M + H)⁺ 442.1879, found 442.1879

(6aS,9aR)-5,6a,7,9a-Tetrahydro-5-phenyl-1-(phenylmethyl)cyclopent[4.5]imidazo[1,2-a]pyrazolo[4,3-e]pyrimidin-4(1H)one (22). Compound 20 (31 mg, 0.07 mmol) was dissolved in dry THF (1 mL). In a separate flask, Pd(OAc)₂ (3.1 mg, 0.2 equiv) was mixed with PPh₃ (18.4 mg, 1 equiv) in dry THF (1 mL). The in situ generated Pd(0) was added to the solution of 20, and the mixture was stirred at room temperature for 5 min. 2-tert-Butylimino-2-diethylamino-1,3-dimethylperhydro-1,3,2-diazaphosphorine 21 (41 μ L, 2 equiv) was added to the mixture, and the reaction was stirred at room temperature for 6 h. Another portion of in situ generated Pd(0) (0.2 equiv) was then added. After 1 h, the reaction was complete. It was then quenched with H₂O and extracted with EtOAc. The organic phase was dried with NaSO₄. After filtration and concentration, the residue was purified by column chromatography eluting with hexanes and EtOAc (1:1–1:4) to give 25 mg (94% yield) of an oil: $R_f = 0.14$ (hexanes/EtOAc = 1:4); $[\alpha]_D = -99.7$ (c = 0.75, CHCl₃); ¹H (500 MHz, CDCl₃) δ 2.56–2.61 (m, 1H), 2.74–2.79 (m, 1H), 4.72 (t, J = 7 Hz, 1H), 5.23 (d, J = 8 Hz, 1H), 5.43 (d, J = 17 Hz, 1H), 5.60–5.61 (m, 1H), 5.65 (d, J = 17 Hz, 1H), 6.10-6.11 (m, 1H), 7.14-7.49 (m, 10H), 7.95 (s, 1H); ¹³C (125 MHz, CDCl_3) δ 40.8, 53.3, 66.9, 68.1, 99.7, 125.8, 126.1, 128.4, 128.6, 128.8, 129.3, 129.5, 135.9, 136.0, 137.7, 138.5, 142.7, 151.6, 157.7; IR (CH₂Cl₂, NaCl) 2918, 1702, 1632, 1592, 1548, 1426, 731 cm⁻¹; HRMS (FAB) calcd for C₂₃H₂₀N₅O (M + H)⁺ 382.1668, found 382.1651.

1-Benzyl-6-cyclopentylamino-5-phenyl-1,5-dihydropyrazolo[3,4-d]pyrimidin-4-one (23). A solution of compound **22** (20 mg, 0.05 mmol) in methanol (5 mL) was charged with Pd/C (10%, 6.6 mg). The mixture was stirred under a balloon filled with H₂. The reaction was monitored by TLC. Upon completion of the reaction, the mixture was diluted with EtOAc and filtered through a thin pad of Celite. The solvent was removed, and the residue was purified by column chromatography eluting with hexanes and EtOAc (1:1) to give 9.4 mg (49% yield) of a white solid: mp = 138–140 °C; $R_f = 0.68$ (hexanes/EtOAc = 1:4); ¹H (500 MHz, CDCl₃) δ 1.25–1.33 (m, 2H), 1.56–1.63 (m, 4H), 2.00–2.08 (m, 2H), 4.11 (d, J = 6.5 Hz, 1H), 4.31, 4.32 (td, J = 8.5, 6.5 Hz), 5.44 (s, 2H), 7.28–7.63 (m. 10H), 7.97 (s, 1H); ¹³C (125 MHz, CDCl₃) δ 23.5, 33.0, 50.5, 53.8, 100.0, 127.7, 128.2, 128.6, 129.1, 129.8, 130.5, 134.8, 135.8, 136.8, 152.4, 152.8, 158.2; IR (CH₂Cl₂, NaCl) 3000, 2987, 1554, 1223, 890 cm⁻¹; HRMS (FAB) calcd for C₂₃H₂₄N₅O (M + H)⁺ 386.1981, found 386.1998.

(6aS,9aR)-5,6a,7,8,9,9a-Hexahydro-5-phenyl-1-(phenylmethyl)cyclopent[4.5]imidazo[1,2-a]pyrazolo[4,3-e]pyrimidine-4(1H)-one (24). To a solution of compound 22 (21 mg, 0.055 mmol) in ethanol (1 mL) were added 35% NH₂NH₂ (0.05 mL, 0.55 mmol) and 30% H₂O₂ (0.07 mL, 0.66 mmol) three times at 8 h intervals. The reaction was quenched with 1 M Na₂S₂O₃ solution, extracted with CH₂Cl₂, washed with brine, and dried with NaSO₄. After filtration and concentration, the residue was purified by column chromatography eluting with hexanes and EtOAc (1:4) to give 10 mg (50% yield) of an oil: $[\alpha]_D = -162$ (c $= 1.0, CHCl_3$; ¹H (500 MHz, CDCl₃) $\delta 1.61-2.09$ (m, 6H), 4.62-4.64 (m, 2H), 5.38 (d, J= 16.5 Hz, 1H), 5.64 (d, J= 17 Hz, 1H), 7.14–7.52 (m, 10H), 7.97 (s, 1H); $^{13}\mathrm{C}$ (125 MHz, CDCl₃) δ 23.1, 34.3, 35.1, 53.3, 62.6, 70.7, 99.7, 126.0, 128.4, 128.6, 128.7, 129.2, $129.5, 136.1, 136.2, 138.5, 142.5, 152.3, 157.7; IR (CH_2Cl_2, NaCl)$ 2960, 1701, 1633, 1591, 1549, 760 cm⁻¹; HRMS (FAB) calcd for $C_{23}H_{22}N_5O (M + H)^+$ 384.1824, found 384.1822.

Acetic Acid 4-(1-Benzyl-4-oxo-5-phenyl-4,5-dihydro-1*H*pyrazolo[3,4-*d*]pyrimidinyl-6-amino)cyclopent-2-enyl Ester (26, *ent*-20). The compound was prepared using the same procedure employed for the synthesis of its enantiomer (20): $[\alpha]_D$ = +8.1 (*c* = 1.0, CHCl₃); ¹H and ¹³C data were identical to those of its enantiomer 20; HRMS (FAB) calcd for C₂₅H₂₄N₅O₃ (M + H)⁺ 442.1879, found 442.1852.

(6a*R*,9a*S*)-5,6a,7,9a-Tetrahydro-5-phenyl-1-(phenylmethyl)cyclopent[4,5]imidazo[1,2-*a*]pyrazolo[4,3-*e*]pyrimidin-4(1*H*)-one (27). The compound was prepared using the same procedure employed for the synthesis of its enantiomer (22): $[\alpha]_D = +100 \ (c = 1.0, \text{ CHCl}_3); R_f = 0.14 \ (\text{hexanes/EtOAc} = 1:4);$ ¹H and ¹³C data were identical to its enantiomer 22; HRMS (FAB) calcd for C₂₃H₂₀N₅O (M + H)⁺ 382.1668, found 382.1659.

(6a*R*,9a*S*)-5,6a,7,8,9,9a-Hexahydro-5-phenyl-1-(phenylmethyl)cyclopent[4,5]imidazo[1,2-*a*]pyrazolo[4,3-*e*]pyrimidin-4(1*H*)one (28). The compound was prepared using the same procedure employed for the synthesis of its enantiomer (24): $[\alpha]_D = +169$ (*c* = 0.93, CHCl₃); $R_f = 0.14$ (hexanes/EtOAc = 1:4); ¹H and ¹³C data were identical to its enantiomer 24; HRMS (FAB) calcd for C₂₃H₂₂N₅O (M + H)⁺ 384.1824, found 384.1830.

Acknowledgment. We thank Eli Lilly and Co. and the NIH for the support of this research. Biological assays were performed by MDS Pharma Services according to literature procedures.¹⁷ We acknowledge Dr. Jaroslav Zajicek for NMR assistance as well as Dr. William Boggess and Nonka Sevova for mass spectroscopy. Special thanks are extended to Maureen Metcalf for her assistance with this manuscript.

Note Added after ASAP Publication. The compound numbers were omitted from the Table of Contents graphic in the version published ASAP March 4, 2005. The corrected version was published March 10, 2005.

Supporting Information Available: ¹H and ¹³C NMR of compounds **20**, **22**, **23**, and **24**. This material is available free of charge via the Internet at http://pubs.acs.org.

JO0484070